

Recent Approaches on the Application of Agro Waste Derived Biocomposites as Green Support Matrix for Enzyme Immobilization

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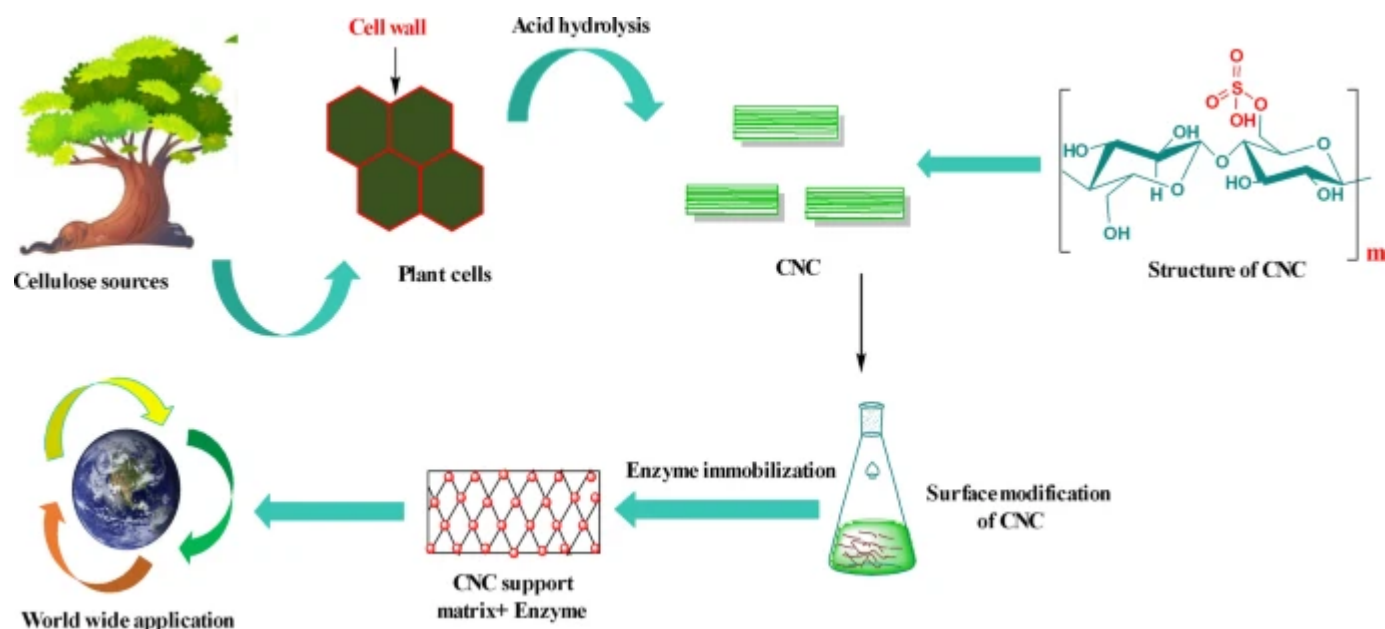
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Abstract

The present imperative interest is in recycling lignocellulosic wastes arising from the global activities of agriculture and forest-based industries. Lignocellulosic waste valorization may help to produce natural biopolymer-based products and their applications in biomedical, biopharmaceutical, and industrial fields. Among these, crystalline nanocellulose (CNC), a cellulose nano-derivative having better enzymatic performance, production yield through immobilization contributes to a green technology and sustainable sources. This is a comprehensive review on the development of cellulose based support matrices, where surface modification of nanocellulose, nanocellulose-protein interactions and applications of cellulose based composites have been illustrated.

Hence, this novel step to recycle primary and secondary agricultural solid waste management will develop pollution free environment, double farmer's income, and cheaper biomaterial for enzyme immobilization that will minimize the process cost value.

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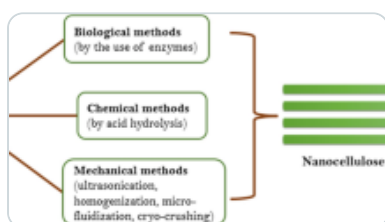
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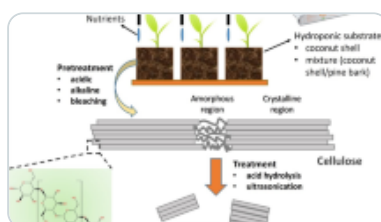
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References

1. APEDA (2014) Agricultural and processed food products export development authority. India New Delhi,
2. Department SR (2020) Production volume of pulp and fruit juice across India from FY 2015 to FY 2020 (in 1000 metric tons). India New Delhi. Accessed 04/09/2020 2020
3. Technology I-CIoP-HE (2020). <https://www.ciphet.in/>. Accessed 04.07.2020 2020
4. Gowman AC, Picard MC, Lim L, Misra M, Mohanty AK (2019) Fruit waste valorization for biodegradable biocomposite applications: a review. BioResources 14(4):10047–10092

[Article](#) [Google Scholar](#)

5. Arsalan A, Younus H (2018) Enzymes and nanoparticles: modulation of enzymatic activity via nanoparticles. Int J Biol Macromol 118:1833–1847

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

6. Nguyen HH, Kim M (2017) An overview of techniques in enzyme immobilization. Appl Sci Converg Technol 26(6):157–163

[Article](#) [Google Scholar](#)

7. Mohamad NR, Marzuki NHC, Buang NA, Huyop F, Wahab RA (2015) An overview of technologies for immobilization of enzymes and surface analysis techniques for immobilized enzymes. Biotechnol Biotechnol Equip 29(2):205–220

[Article](#) [PubMed](#) [PubMed Central](#) [CAS](#) [Google Scholar](#)

8. Zhang B, Weng Y, Xu H, Mao Z (2012) Enzyme immobilization for biodiesel production. Appl Microbial Biotechnol 93(1):61–70

9. Sirisha VL, Jain A, Jain A (2016) Enzyme immobilization: an overview on methods, support material, and applications of immobilized enzymes. *Adv in Food and Nutr Res* 79:179–211

[Article](#) [CAS](#) [Google Scholar](#)

10. Zhang H, Lyu S, Zhou X, Gu H, Ma C, Wang C, Ding T, Shao Q, Liu H, Guo Z (2019) Super light 3D hierarchical nanocellulose aerogel foam with superior oil adsorption. *J Colloid Interface Sci* 536:245–251

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

11. Ahmed IN, Yang X-L, Dubale AA, Li R-F, Ma Y-M, Wang L-M, Hou G-H, Guan R-F, Xie M-H (2018) Hydrolysis of cellulose using cellulase physically immobilized on highly stable zirconium based metal-organic frameworks. *Bioresour Technol* 270:377–382

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

12. Ali M, Husain Q, Sultana S, Ahmad M (2018) Immobilization of peroxidase on polypyrrole-cellulose-graphene oxide nanocomposite via non-covalent interactions for the degradation of Reactive Blue 4 dye. *Chemosphere* 202:198–207

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

13. Yakout AA, El-Sokkary RH, Shreadah MA, Hamid OGA (2017) Cross-linked graphene oxide sheets via modified extracted cellulose with high metal adsorption. *Carbohydr Polym* 172:20–27

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

14. Xu C, Molino BZ, Wang X, Cheng F, Xu W, Molino P, Bacher M, Su D, Rosenau T, Willför S (2018) 3D printing of nanocellulose hydrogel scaffolds with tunable mechanical strength towards wound healing application. *J Mater Chem B* 6(43):7066–7075

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

15. Zhao X, Qi F, Yuan C, Du W, Liu D (2015) Lipase-catalyzed process for biodiesel production: enzyme immobilization, process simulation and optimization. *Renew Sust Energ Rev* 44:182–197

[Article](#) [CAS](#) [Google Scholar](#)

16. Ghodake GS, Yang J, Shinde SS, Mistry BM, Kim D-Y, Sung J-S, Kadam AA (2018) Paper waste extracted α -cellulose fibers super-magnetized and chitosan-functionalized for covalent laccase immobilization. *Bioresour Technol* 261:420–427

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

17. Soltani Firooz N, Panahi R, Mokhtarani B, Yazdani F (2017) Direct introduction of amine groups into cellulosic paper for covalent immobilization of tyrosinase: support characterization and enzyme properties. *Cellulose* 24:1407–1416

[Article](#) [CAS](#) [Google Scholar](#)

18. Vasconcelos NF, Andrade FK, Vieira LdAP, Vieira RS, Vaz JM, Chevallier P, Mantovani D, de Fátima BM, de Freitas RM (2020) Oxidized bacterial cellulose membrane as support for enzyme immobilization: properties and morphological features. *Cellulose* 27(6):3055–3083

[Article](#) [CAS](#) [Google Scholar](#)

19. Wang L, Zhi W, Wan J, Han J, Li C, Wang Y (2019) Recyclable β -glucosidase by one-pot encapsulation with Cu-MOFs for enhanced hydrolysis of cellulose to glucose. *ACS Sustain Chem Eng* 7(3):3339–3348

[Article](#) [CAS](#) [Google Scholar](#)

20. Patel SR, Yap MGS, Wang DIC (2009) Immobilization of l-lactate dehydrogenase on magnetic nanoclusters for chiral synthesis of pharmaceutical compounds. *Biochem Eng J* 48(1):13–21

[Article](#) [Google Scholar](#)

21. Prakasham RS, Devi GS, Rao CS, Sivakumar V, Sathish T, Sarma PN (2010) Nickel-impregnated silica nanoparticle synthesis and their evaluation for biocatalyst immobilization. *Appl Biochem Biotechnol* 160(7):1888–1895

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

22. Singh P, Magalhães S, Alves L, Antunes F, Miguel M, Lindman B, Medronho B (2019) Cellulose-based edible films for probiotic entrapment. *Food Hydrocoll* 88:68–74

[Article](#) [CAS](#) [Google Scholar](#)

23. Qiao L, Li S, Du K (2020) Fabrication and characterization of porous cellulose beads with high strength and specific surface area via preliminary chemical cross-linking reaction for protein separation. *Biochem Eng J* 153:107412

[Article](#) [CAS](#) [Google Scholar](#)

24. de Mello DC (2014) Nanoparticles: workhorses of nanoscience. Springer, Berlin

[Google Scholar](#)

25. Ealia SAM, Saravanakumar M A review on the classification, characterisation, synthesis of nanoparticles and their application. In: IOP Conference Series: Materials Science and Engineering, 2017. vol 3. IOP Publishing, p 032019
26. Kruger CA, Abrahamse H (2018) Utilisation of targeted nanoparticle photosensitiser drug delivery systems for the enhancement of photodynamic therapy. *Molecules* 23(10):2628

[Article](#) [PubMed Central](#) [Google Scholar](#)

27. Ding S, Cargill AA, Medintz IL, Claussen JC (2015) Increasing the activity of immobilized enzymes with nanoparticle conjugation. *Curr Opin Biotechnol* 34:242–250

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

28. Bilal M, Iqbal HM (2019) Naturally-derived biopolymers: Potential platforms for enzyme immobilization. *Int J Biol Macromol* 130:462–482

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

29. Grigoros AG (2017) Catalase immobilization—a review. *Biochem Eng J* 117:1–20

[Article](#) [CAS](#) [Google Scholar](#)

30. de Martins OS, Moreno-Perez S, Romero-Fernández M, Fernandez-Lorente G, Rocha-Martin J, Guisan JM (2018) Immobilization and stabilization of commercial β -1, 4-endoxylanase DepolTM 333MDP by multipoint covalent attachment for xylan hydrolysis: Production of prebiotics (xylo-oligosaccharides). *Biocatal Biotransfor* 36(2):141–150

[Article](#) [Google Scholar](#)

31. Lokha Y, Arana-Peña S, Rios NS, Mendez-Sanchez C, Gonçalves LR, Lopez-Gallego F, Fernandez-Lafuente R (2020) Modulating the properties of the lipase from *Thermomyces lanuginosus* immobilized on octyl agarose beads by altering the immobilization conditions. *Enzyme Microb Technol* 133:109461

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

32. Karim A, Bibi Z, Rehman HU, Aman A, Qader SAU, Rashid MH (2021) Single step immobilization of CMCase within agarose gel matrix: Kinetics and thermodynamic studies. *Colloids Surf B* 200:111583

[Article](#) [CAS](#) [Google Scholar](#)

33. Serey M, Vera C, Guerrero C, Illanes A (2021) Immobilization of *Aspergillus oryzae* β -galactosidase in cation functionalized agarose matrix and its application in the synthesis of lactulose. *Int J Biol Macromol* 167:1564–1574

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

34. Lopes LA, Novelli PK, Fernandez-Lafuente R, Tardioli PW, Giordano RLC (2020) Glyoxyl-activated agarose as support for covalently link Novo-Pro D: biocatalysts performance in the hydrolysis of casein. *Catalysts* 10(5):466

[Article](#) [CAS](#) [Google Scholar](#)

35. Garcia-Bofill M, Sutton PW, Guillen M, Alvaro G (2021) Enzymatic synthesis of a statin precursor by immobilised alcohol dehydrogenase with NADPH oxidase as cofactor regeneration system. *Appl Cat A* 609:117909

[Article](#) [CAS](#) [Google Scholar](#)

36. Kamaci UD, Peksel A (2021) Enhanced catalytic activity of immobilized phytase into polyvinyl alcohol-sodium alginate based electrospun nanofibers. *Cat Lett* 151(3):821–831

37. Yavaşer R, Karagözler AA (2021) Laccase immobilized polyacrylamide–alginate cryogel: a candidate for treatment of effluents. *Process Biochem* 101:137–146

38. Guo R, Zheng X, Wang Y, Yang Y, Ma Y, Zou D, Liu Y (2021) Optimization of cellulase immobilization with sodium alginate–polyethylene for enhancement of enzymatic hydrolysis of microcrystalline cellulose using response surface methodology. *Appl Biochem Biotechnol* 193:1–18

39. Marjani A, Zare MH, Sadeghi MH, Shirazian S, Ghadiri M (2021) Synthesis of alginate–coated magnetic nanocatalyst containing high–performance integrated enzyme for phenol removal. *J Environ Chem Eng* 9(1):104884

40. Qamar SA, Asgher M, Bilal M (2020) Immobilization of alkaline protease from *Bacillus brevis* using Ca–alginate entrapment strategy for improved catalytic stability, silver recovery, and dehairing potentialities. *Cat Lett* 150:3572–3583

41. Maity M, Bhattacharyya A, Bhowal J (2021) Production and immobilization of β –galactosidase isolated from *Enterobacter aerogenes* KCTC2190 by entrapment method using agar–agar organic matrix. *J Appl Biochem Biotechnol*. 193:1–27

42. Irfan M, Kiran J, Ayubi S, Ullah A, Rana QUA, Khan S, Hasan F, Badshah M, Shah AA (2020) Immobilization of β –1, 4–xylanase isolated from *Bacillus licheniformis* S3. *J*

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

43. Kalita T, Sangma SD, Bez G, Ambasht P (2020) Immobilization of acid phosphatase in agar-agar and gelatin: comparative characterization. J Sci Res 64(2):192–200

[Google Scholar](#)

44. Martín MC, DellÓlio GAG, Villar MA, Morata VI, López OV, Ninago MD Preparation and Characterization of an Immobilized Enological Pectinase on Agar-Alginate Beads. In: Macromolecular Symposia, 2020. vol 1. Wiley Online Library, p 1900208

45. Pervez S, Nawaz MA, Jamal M, Jan T, Maqbool F, Shah I, Aman A, Qader SAU (2019) Improvement of catalytic properties of starch hydrolyzing fungal amyloglucosidase: utilization of agar-agar as an organic matrix for immobilization. Carbohydr Res 486:107860

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

46. Tavares L, Flores EEE, Rodrigues RC, Hertz PF, Noreña CPZ (2020) Effect of deacetylation degree of chitosan on rheological properties and physical chemical characteristics of genipin-crosslinked chitosan beads. Food Hydrocoll 106:105876

[Article](#) [CAS](#) [Google Scholar](#)

47. Ji S, Liu W, Su S, Gan C, Jia C (2021) Chitosan derivative functionalized carbon nanotubes as carriers for enzyme immobilization to improve synthetic efficiency of ethyl caproate. LWT 149:111897

[Article](#) [CAS](#) [Google Scholar](#)

48. Srivastava B, Singh H, Khatri M, Singh G, Arya SK (2020) Immobilization of keratinase on chitosan grafted- β -cyclodextrin for the improvement of the enzyme

properties and application of free keratinase in the textile industry. *Int J Biol Macromol* 165:1099–1110

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

49. Lima PC, Gazoni I, de Carvalho AMG, Bresolin D, Cavalheiro D, de Oliveira D, Rigo E (2021) β -galactosidase from *Kluyveromyces lactis* in genipin-activated chitosan: An investigation on immobilization, stability, and application in diluted UHT milk. *Food Chem* 349:129050

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

50. Ricardi NC, Arenas LT, Benvenutti EV, Hinrichs R, Flores EEE, Hertz PF, Costa TMH (2021) High performance biocatalyst based on β -d-galactosidase immobilized on mesoporous silica/titania/chitosan material. *J Food Chem* 359:129890

[Article](#) [CAS](#) [Google Scholar](#)

51. Teepoo S, Dawan P, Barnthip N (2017) Electrospun chitosan-gelatin biopolymer composite nanofibers for horseradish peroxidase immobilization in a hydrogen peroxide biosensor. *J Biosensors* 7(4):47

[Article](#) [Google Scholar](#)

52. Nishida VS, de Oliveira RF, Brugnari T, Correa RCG, Peralta RA, Castoldi R, de Souza CGM, Bracht A, Peralta RM (2018) Immobilization of *Aspergillus awamori* β -glucosidase on commercial gelatin: an inexpensive and efficient process. *Int J Biol Macromol* 111:1206–1213

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

53. Asgher M, Noreen S, Bilal M (2017) Enhancement of catalytic, reusability, and long-term stability features of *Trametes versicolor* IBL-04 laccase immobilized on different polymers. *Int J Biol Macromol* 95:54–62

54. Zhang J, Ke W, Chen H (2020) Enhancing laccase production by white-rot fungus *trametes hirsuta* SSM-3 in co-culture with yeast *sporidiobolus pararoseus* SSM-8. *Prep Biochem Biotechnol* 50(1):10–17

55. Awad GEA, Ghanem AF, Wahab WAA, Wahba MI (2020) Functionalized κ -carrageenan/hyperbranched poly (amidoamine) for protease immobilization: thermodynamics and stability studies. *Int J Biol Macromol* 148:1140–1155

56. Wahba MI, Hassan ME, Ali KA (2021) Chitosan–glutaraldehyde activated carrageenan–alginate beads for β -D-galactosidase covalent immobilisation. *J Biocat Biotransform* 39(2):138–151

57. Ali AO, Abdalla MS, Shahein YE, Shokeer A, Sharada HM, Ali KA (2021) Grafted carrageenan: alginate gel beads for catalase enzyme covalent immobilization. *3 Biotech*. 11(7):1–7

58. Carneiro EA, Bastos AKP, Oliveira UMFd, Matos LJBLd, Adriano WS, Monteiro RRdC, Santos JCSd, Gonçalves LRB (2020) Improving the catalytic features of the lipase from *Rhizomucor miehei* immobilized on chitosan-based hybrid matrices by altering the chemical activation conditions. *J Química Nova* 43:1234–1239

59. Sannino A, Demitri C, Madaghiele M (2009) Biodegradable cellulose-based hydrogels: design and applications. *Materials* 2(2):353–373

[Article](#) [PubMed Central](#) [CAS](#) [Google Scholar](#)

60. Nikolic T, Korica M, Milanovic J, Kramar A, Petronijevic Z, Kostic M (2017) TEMPO-oxidized cotton as a substrate for trypsin immobilization: impact of functional groups on proteolytic activity and stability. *Cellulose* 24(4):1863–1875

[Article](#) [CAS](#) [Google Scholar](#)

61. Talingtaisong S, Vongsetskul T, Panatdasirisuk W, Tangboriboonrat P (2017) Gauze-reinforced electrospun regenerated cellulose ultrafine fibers for immobilizing bromelain. *Cellulose* 24(7):2967–2975

[Article](#) [CAS](#) [Google Scholar](#)

62. Feng Q, Hou D, Zhao Y, Xu T, Menkhaus TJ, Fong H (2014) Electrospun regenerated cellulose nanofibrous membranes surface-grafted with polymer chains/brushes via the atom transfer radical polymerization method for catalase immobilization. *ACS Appl Mater Interfaces* 6(23):20958–20967

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

63. Andrés-Bello A, Iborra-Bernad C, García-Segovia P, Martínez-Monzó J (2013) Effect of konjac glucomannan (KGM) and carboxymethylcellulose (CMC) on some physico-chemical and mechanical properties of restructured gilthead sea bream (*Sparus aurata*) products. *Food Bioproc Tech* 6(1):133–145

[Article](#) [Google Scholar](#)

64. Liu S, Zhu Y, Li W, Li Y, Li B (2016) Preparation of a magnetic responsive immobilized lipase–cellulose microgel catalyst system: role of the surface properties of the magnetic cellulose microgel. *RSC Adv* 6(9):7339–7347

65. Sillu D, Agnihotri S (2019) Cellulase immobilization onto magnetic halloysite nanotubes: enhanced enzyme activity and stability with high cellulose saccharification. *ACS Sustain Chem Eng* 8(2):900–913

[Article](#) [Google Scholar](#)

66. Rajdeo K, Harini T, Lavanya K, Fadnavis NW (2016) Immobilization of pectinase on reusable polymer support for clarification of apple juice. *Food Bioprod Process* 99:12–19

[Article](#) [CAS](#) [Google Scholar](#)

67. Bilal M, Iqbal HMN (2019) Sustainable bioconversion of food waste into high-value products by immobilized enzymes to meet bio-economy challenges and opportunities—A review. *Food Res Int* 123:226–240

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

68. Wu Z, Deng W, Luo J, Deng D (2019) Multifunctional nano-cellulose composite films with grape seed extracts and immobilized silver nanoparticles. *Carbohydr polym* 205:447–455

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

69. Shak KPY, Pang YL, Mah SK (2018) Nanocellulose: recent advances and its prospects in environmental remediation. *Beilstein J Nanotechnol* 9:2479–2498

[Article](#) [PubMed](#) [PubMed Central](#) [CAS](#) [Google Scholar](#)

70. Phanthong P, Reubroycharoen P, Hao X, Xu G, Abudula A, Guan G (2018) Nanocellulose: extraction and application. *Carbon Resour Convers* 1(1):32–43

71. Kaushik M, Moores A (2016) Nanocelluloses as versatile supports for metal nanoparticles and their applications in catalysis. *Green Chem* 18(3):622–637

[Article](#) [CAS](#) [Google Scholar](#)

72. Markets MA (2020) Global Forecast to 2025.
<https://www.marketsandmarkets.com/pdfdownloadNew.asp?id=56392090>.
Accessed 04.03.2020 2020

73. Sharma A, Rana H, Goswami S (2021) A comprehensive review on the heavy metal removal for water remediation by the application of lignocellulosic biomass-derived nanocellulose. *J Polym Environ* 30:1–18

[Article](#) [Google Scholar](#)

74. Wahab RA, Elias N, Abdullah F, Ghoshal SK (2020) On the taught new tricks of enzymes immobilization: an all-inclusive overview. *React Funct Polym* 152:104613

[Article](#) [CAS](#) [Google Scholar](#)

75. Steen RE, Ta DT, Cortens D, Billen B, Guedens W, Adriaensens P (2013) Protein engineering for directed immobilization. *Bioconjugate Chem* 24(11):1761–1777

[Article](#) [Google Scholar](#)

76. Abushammala H, Mao J (2019) A review of the surface modification of cellulose and nanocellulose using aliphatic and aromatic mono- and di-isocyanates. *Molecules* 24(15):2782

[Article](#) [PubMed Central](#) [Google Scholar](#)

77. Vaz Graaf M, De LA, Reis RL (2003) In vitro degradation behaviour of biodegradable soy plastics: effects of crosslinking with glyoxal and thermal treatment. Polym Degradatln Stabil. 81(65):74

[Google Scholar](#)

78. Altun GD, Cetinus SA (2007) Immobilization of pepsin on chitosan beads. Food Chem 100(964):971

[Google Scholar](#)

79. Bindu VU, Mohanan PV (2020) Thermal deactivation of α -amylase immobilized magnetic chitosan and its modified forms : a kinetic and thermodynamic study. Carbohydr Res 498:108185

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

80. Bezerra CS, Maria Farias C, De Sousa G, De M (2015) Enzyme immobilization onto renewable polymeric matrixes : past, present, and future trends. J Appl polym 42125(1):15. <https://doi.org/10.1002/app.42125>

[Article](#) [CAS](#) [Google Scholar](#)

81. Rodríguez-Restrepo YA, Orrego CE (2020) Immobilization of enzymes and cells on lignocellulosic materials. Environ Chem Lett 18(3):787–806

[Article](#) [Google Scholar](#)

82. Kontturi E, Laaksonen P, Linder MB, Gröschel AH, Rojas OJ, Ikkala O (2018) Advanced materials through assembly of nanocelluloses. Adv Mater 30(24):1703779

[Article](#) [Google Scholar](#)

83. Incani V, Danumah C, Boluk Y (2013) Nanocomposites of nanocrystalline cellulose for enzyme immobilization. *Cellulose* 20(1):191–200

[Article](#) [CAS](#) [Google Scholar](#)

84. Eibinger M, Sattelkow J, Ganner T, Plank H, Nidetzky B (2017) Single-molecule study of oxidative enzymatic deconstruction of cellulose. *Nat commun* 8(1):1–7

[Article](#) [CAS](#) [Google Scholar](#)

85. Liu Y, Chen JY (2016) Enzyme immobilization on cellulose matrixes. *J Bioact Compat Polym* 31(6):553–567

[Article](#) [CAS](#) [Google Scholar](#)

86. Costa-Silva TA, Souza CRE, Said S, Oliveira WP (2015) Drying of enzyme immobilized on eco-friendly supports. *Afr J Biotechnol* 14(44):3019–3026

[Article](#) [CAS](#) [Google Scholar](#)

87. Sunasee R, Hemraz UD, Ckless K (2016) Cellulose nanocrystals: a versatile nanoplatform for emerging biomedical applications. *Expert Opin Drug Deliv* 13(9):1243–1256

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

88. Uth C, Zielonka S, Hörner S, Rasche N, Plog A, Orelma H, Avrutina O, Zhang K, Kolmar H (2014) A chemoenzymatic approach to protein immobilization onto crystalline cellulose nanoscaffolds. *Angew Chem Int Ed* 53(46):12618–12623

[CAS](#) [Google Scholar](#)

89. Schyrr B, Pasche S, Voirin G, Weder C, Simon YC, Foster EJ (2014) Biosensors based on porous cellulose nanocrystal–poly (vinyl alcohol) scaffolds. *ACS Appl Mater Interfaces* 6(15):12674–12683

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

90. Valo H, Arola S, Laaksonen P, Torkkeli M, Peltonen L, Linder MB, Serimaa R, Kuga S, Hirvonen J, Laaksonen T (2013) Drug release from nanoparticles embedded in four different nanofibrillar cellulose aerogels. *Eur J Pharm Sci* 50(1):69–77

[Article](#) [PubMed](#) [CAS](#) [Google Scholar](#)

91. Fritz C, Jeuck B, Salas C, Gonzalez R, Jameel H, Rojas OJ (2015) Nanocellulose and proteins: exploiting their interactions for production, immobilization, and synthesis of biocompatible materials. *Cellul Chem Prop.* 271(207):224

[Google Scholar](#)

92. Fu LH, Qi C, Liu YJ, Cao WT, Ma MG (2018) Sonochemical synthesis of cellulose/hydroxyapatite nanocomposites and their application in protein adsorption. *Sci Rep* 8(1):1–12

[Article](#) [Google Scholar](#)

93. Lim EK, Chung BH (2016) Preparation of pyrenyl-based multifunctional nanocomposites for biomedical applications. *Nat prot* 11(2):236–251

[Article](#) [CAS](#) [Google Scholar](#)

94. Alonso-Lerma B, Barandiaran L, Ugarte L, Larraza I, Reifs A, Olmos-Juste R, Barruetabeña N, Amenabar I, Hillenbrand R, Eceiza A (2020) High performance crystalline nanocellulose using an ancestral endoglucanase. *Commun Mater* 1(1):1–10

95. Rosa L, Blackledge J, Boretti A (2017) Nano-magnetic resonance imaging (nano-MRI) gives personalized medicine a new perspective. *Biomedicines* 5(1):7

[Article](#) [PubMed Central](#) [Google Scholar](#)

96. Petropoulou A, Kralj S, Karagiorgis X, Savva I, Loizides E, Panagi M, Krasia-Christoforou T, Riziotis C (2020) Multifunctional gas and pH fluorescent sensors based on cellulose acetate electrospun fibers decorated with rhodamine B-functionalised core-shell ferrous nanoparticles. *Sci rep* 1(1):1–14

[Google Scholar](#)

97. Lin X, Han X, Wang J (2018) In situ synthesis of easily separable Au nanoparticles catalysts based on cellulose hydrogels. *Polym J* 50(7):495–501

[Article](#) [CAS](#) [Google Scholar](#)

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Contributions

All authors contributed to the study conception and design. HR and SD: Material preparation, data collection and analysis were performed. The first draft of the manuscript was written by HR and all authors commented on previous versions of the manuscript. All authors read and approved the final manuscript.

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Ethics declarations

Conflict of interest

Authors have no conflict of interest to disclose in any capacity either competing or financial.

Additional information

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